Note

Structures and amounts of branched molecules in rice amyloses

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Rice starches with low (japonica varieties) and high (indica varieties) affinities for iodine have similar contents of amylose but different affinities of their amylopectins for iodine¹. Their amyloses have the same affinity for iodine and show no significant differences in molecular size, average chain-length, and number of chains². However, the structures and amounts of their branched molecules have not been examined except for the amylose from a japonica, Sasanishiki variety³. We now report on five rice amyloses from starches with low [japonica: Hokkaido (unknown variety, produced in Hokkaido), and Sasanishiki] and high (indica: IR32, IR36, and IR42) affinities for iodine.

No method is available at present for the isolation of branched amyloses. Therefore, the beta-limit dextrins (β -LD) were isolated³ after treatment of the amylose fraction with beta-amylase in order to remove all the linear molecules, and their properties are recorded in Table I. The iodine staining and binding properties of each β -LD were similar to those of the parent amylose, and their number- $(\overline{d.p.}_{n})$ and weight-average $(\overline{d.p.}_{w})$ degrees of polymerization were similar to each other except for IR32 β -LD, for which the values were slightly lower. These values were higher than those of amylomaize β -LD (d.p., 690–710, d.p., 1910–1990)⁴, similar to those of corn β -LD ($\overline{\text{d.p.}}_n$ 790–850, $\overline{\text{d.p.}}_w$ 2700–3000)⁵, and lower than those of water chestnut $(\overline{d.p.}_n 1200, \overline{d.p.}_w 4150)^6$ and sago $(\overline{d.p.}_n 1830-4090, \overline{d.p.}_w 4150 (12,500)^7$ β -LDs. The rice β -LDs had similar molecular sizes, close to those (d.p., 920-1110, d.p., 2480-3500) of their respective parent amyloses and to those of corn⁵, amylomaize⁴, and sago amyloses⁷. The average chain-lengths (c.l.) of the rice β-LDs were in the range 90-155, but the IR36 β-LD had a slightly higher c.l. This range resembles that for amyloses from other plants³⁻⁶ and suggests that the branched molecules from rice have a similar inner chain-length. The average

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TABLE I				
PROPERTIES	OF BETA-LIMIT	DEXTRINS E	ROM RICE	AMVLOSES

Property	Japonica		Indica		
	Hokkaido	Sasanishiki	IR32	IR36	IR42
Iodine affinity (g/100 g)	18.8	19.5a	19.4	19.9	18.5
Blue value	1.35	1.35^{a}	1.37	1.40	1.35
$\frac{\lambda_{\mathrm{max}}}{(\mathrm{nm})}$	653	659^{a}	650	660	653
$\overline{\mathbf{D}.\mathbf{p}{\mathbf{n}}^{h}}$	890	1030^{a}	790	890	840
D.p	3080	3500	2480	3460	2760
D.p. _n /d.p. _w	3.46	3.40	3.10	3.89	3.39
Apparent d.p. distribution ^d	440-10,200	420-11,500	460-7400	410-11,300	370-9200
Average chain-length (c.l.)	105	115^{a}	105	155	90
Number of chains	8.5	9.0^a	7.5	5.7	9.7

^aFrom ref. 3. ^{b.c}Number- and weight-average degrees of polymerization, respectively. ^d \overline{D} .p. values of the sub-fractions (10% by weight) having the lowest and highest molecular weights.

number of chains of the β -LDs indicate that the branched molecule from IR36 had 5.7 chains on average, whereas the others had 7.5–9.7 chains. These numbers were similar to those of branched molecules from most other plants except those from tapioca (17.1) and sago (17.3 and 19.4)^{3–6}.

Fig. 1 shows h.p.l.c. gel-permeation chromatograms of the β -LDs. Each β -LD gave a single peak with a maximum in the range d.p. 2240-3120. The elution profile resembled that of the parent amylose². The linear plots of d.p. against retention time imply that both large and small β -LDs have similar hydrodynamic properties, due to their similar shapes. The β -LDs from water chestnut⁶ and sago⁷ have been suggested to be large and more spherical in shape, with more extensive branching, because of their concave curves of d.p. versus retention time in gelpermeation chromatography. The apparent distributions of d.p. of most rice β -LDs were in the range of 400–12,000, and the IR32 β -LD had a narrower distribution (460–7400). A similar range (200–13,000) was reported² for their parent amyloses. The high $\overline{d.p.}_{w}/\overline{d.p.}_{n}$ values (3.1-3.9) of the β -LDs suggest that the branched molecules from rice have a little broader distribution of molecular weights than their parent amyloses² (2.6–3.4). These resemblances between the β -LDs and their parent amyloses may imply that the branched molecules from rice are a little larger than the unbranched molecules and are degraded to small extents with betaamylase, but have distributions of d.p. similar to those of the unbranched molecules.

Thus, the β -LDs have similar structures and, on the assumption that the branched molecules have a similar beta-amylolysis limit, there seem to be no significant differences.

The molar fractions (Table II) of the branched molecules from rice except that (0.49) of the IR32 molecule were $\sim 0.3-0.4$, similar to those of cereal³, nut^{3,6},

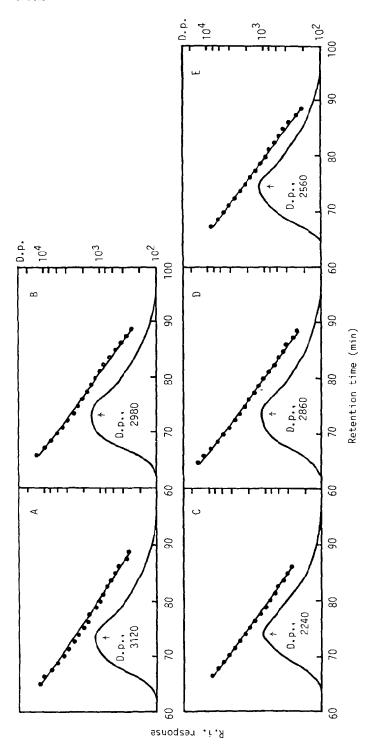


Fig. 1. H.p.l.c. elution curves for beta-limit dextrins from rice amyloses (A, B, C, D, and E: Hokkaido, Sasanishiki, IR32, IR36, and IR42 varieties, respectively). The conditions were as reported¹¹. —, Response of the differential refractometer (R.i.). • D.p.

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TABLE II	
MOLAR EPACTIONS" OF REANCHED AND LINRRANCHED MOLECULES IN R	ICE AMYLOSES

	Japonica		Indica		
	Hokkaido	Sasanishiki	IR32	IR36	IR42
Branched molecule	0.43	0.31	0.49	0.32	0.38
Unbranched molecule	0.57	0.69	0.51	0.68	0.62

"Molar fractions (Mf) were calculated from the average numbers of chains (Nc) of amylose and its beta-limit dextrin by the following equations: Mf of branched molecule = $(Nc_{amylose} - 1)/(Nc_{limit dextrin} - 1)$, Mf of unbranched molecule = $1 - Mf_{branched molecule}$.

root, tuber³, and trunk⁷ amyloses but different from that (0.70) of sweet-potato amylose³, suggesting that the branched molecules are the minor component in most rice amyloses.

EXPERIMENTAL

Materials. — Pure rice amyloses from japonica and indica varieties were those used previously². β -LDs from the amyloses were prepared as described³.

Analytical methods. — Iodine affinity was determined at 25° by a modified¹ amperometric titration8. Blue values were determined as described9. The d.p.n and c.l. were determined by the modified Park–Johnson method and the rapid Smith-degradation method¹0 with minor modifications², respectively. The number of chains per molecule was calculated as d.p.n/c.l. The d.p.w and apparent distribution of d.p. were determined by gel-permeation h.p.l.c., using connected columns (Tosoh TSKgel G6000PW, G4000PW, and G3000PW) with a differential refractometer (Tosoh RI-8000) and a low-angle laser-light-scattering photometer (Tosoh LS-8) as detectors¹¹.

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